

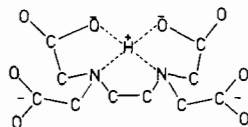
Anomalous H/D Isotopic Substitution Effects in Aqueous Solutions of Ethylenediaminetetra-acetic Acid

R. H. NUTTALL*, D. M. STALKER and I. THOM

Department of Pure and Applied Chemistry, University of Strathclyde, Glasgow, G1 1XL, Scotland

Received January 19, 1978

We have previously reported the Raman spectra of aqueous (H₂O) solutions containing the ions HEDTA³⁻ (pH 8) and H₂EDTA²⁻ (pH 5) (H₄EDTA = ethylenediaminetetra-acetic acid), with Na⁺ as counter ion [1]. The data were interpreted as favouring a folded configuration resulting from the formation of intramolecular hydrogen bonds between the carboxylate groups and protonated nitrogen atoms. The arrangement is analogous to that proposed for the ion HCDTA³⁻ (CDTA = 1,2-cyclohexanediamine-tetra-acetic acid) on the basis of PMR evidence [2].



The initial protonation of EDTA⁴⁻ in H₂O solution is accompanied by a marked increase in the frequency of all the Raman active ν C–H vibrations, apparently indicative of the involvement of both nitrogens in the initial protonation process [1, 3, 4]. The Na-23 NMR spectra [5] of these solutions also demonstrate that the process of monoprotonation completely displaces the Na⁺ ion from its weak interaction with EDTA⁴⁻. In D₂O the conversion of EDTA⁴⁻ to DEDTA³⁻ may be followed on the basis of the changes which occur in the ν_{asym} CO₂⁻ region of the

IR spectrum: thus EDTA⁴⁻ is characterised by a single band at 1580 cm⁻¹ but DEDTA³⁻ gives rise to two bands of similar intensity at 1580 and 1620 cm⁻¹; at pD 5 where D₂EDTA²⁻ is present solely the band at 1620 cm⁻¹ remains [6, 7]. For the H₂O solutions the process of monoprotonation is also accompanied by a shift of the single Raman active ν C–C vibration of the carboxylate arms; diprotonation to H₂EDTA²⁻ causes this band to split into two components [1]. However Krishan and Plane [8], who examined one EDTA/Na⁺/D₂O solution in the course of a Raman study of a range of EDTA containing solutions, reported that at pD 4 only one ν C–C band was detectable as opposed to the two bands observed for the equivalent H₂O solution. In order to confirm and extend this apparently anomalous observation we have examined the Raman spectra of a series of EDTA/Na⁺/D₂O solutions from pD 4 to 12. These spectra show the same shifts of ν C–H with pD as were observed for the H₂O solutions but a different pattern for the ν C–C bands. Thus whereas the spectrum of aqueous HEDTA³⁻ has only one ν C–C band, that of DEDTA³⁻ in D₂O has two; at pD 5 where D₂EDTA²⁻ is the main species only one ν C–C band is present, confirming Krishnan and Plane's result (Table I).

PMR studies have determined [9] that the rate of deuteration of either the ethylenic, or the methylenic, protons in EDTA/Na⁺/D₂O solutions is negligible so that any H/D shifts in the vibrational spectra must be attributed to the formation of N–D⁺ vs. N–H⁺ bonds, or to the influence of interaction with the solvent. In order to ensure that we are indeed comparing like 'monoprotonated' and 'diprotonated' species we have made use of the internally consistent pD scale developed by Paabo and Bates [10] and also obtained the IR spectra of the D₂O solutions. By a comparison with the data obtained by Nakamoto *et al.* [7] it is possible to identify the solution at pD 8.45 as certainly containing the DEDTA³⁻ ion. Unfortunately the strong H₂O band at 1650 cm⁻¹ prevents complete observation of the changes

*Address for correspondence.

TABLE I. ν C–H and ν C–C Frequencies in the Raman Spectra of Na⁺/EDTA/D₂O Solutions (cm⁻¹).

pD	ν C–H	ν C–C ^a
12.60		2956(vs) 2853(vs) 918(s)
9.32		2984(sh) 2956(vs) 2856(s) 916(s, br)
8.45	3028(w)	2987(vs) 2956(w) 2870(w) 923(m) 908(s)
7.43	3020(m)	2976(vs) 2870(w) 926(s) 901(w)
6.30	3020(m)	2976(vs) 925(s, br)
5.0	3018(m)	2976(vs) 930(s, br)
4.32	3068(w) 3022(w)	2985(vs) 930(s, br)

^aAt pH 8.47, in H₂O solution, ν C–C is at 922 cm⁻¹ and at pH 5.05 at 935 and 911 cm⁻¹ [1].

in the IR carboxylate region which take place in H_2O as EDTA^{4-} is converted to HEDTA^{3-} . For the former a shoulder at 1580 cm^{-1} is prominent; with mono-protonation this diminishes in intensity and the water peak increases in intensity, however at pH 8 the shoulder is still detectable. This observation, coupled with the changes in the $\nu\text{C-H}$ region of the Raman spectra which are common to H_2O and D_2O solutions suggests that basically the same process takes place in both H_2O and D_2O . A preliminary examination of CDTA in H_2O and D_2O demonstrates that similar changes in the $\nu\text{C-H}$ region occur with 'protonation', though for this system changes in the $\nu\text{C-C}$ region are more complex and also complicated by the spectrum of the cyclohexane ring; detailed study requires higher resolution than we currently have at our disposal.

Previous studies of glycine and ethylenediamine in $\text{H}_2\text{O}/\text{D}_2\text{O}$ solutions have resulted in assignments of N-H stretching modes above 3000 cm^{-1} and deformation modes at about 1600 cm^{-1} [3, 4]. While these assignments for both N-H and N-D frequencies are complicated by their proximity to the vibrational modes of H_2O and D_2O respectively, there is no evidence to suggest that N-D vibrations should occur at 900 cm^{-1} . It is thus clear that the changes in vibrational pattern which we record are not associated with normal isotopic H/D substitution displacements. Similarly anomalous changes were observed in the Raman spectra of glycine and ethylenediamine in $\text{H}_2\text{O}/\text{D}_2\text{O}$ solutions and it appears that the effects may be specific to the formation of hydrogen and deuterium bonds. It seems likely that the differences at least in $\text{H}_2\text{O}/\text{D}_2\text{O}/\text{EDTA}$ solutions, reflect a

variation in the strain in the rings which are formed as a result of intramolecular hydrogen bonding. Such a result would ensure if, for example, hydrogen and deuterium bonds differed significantly in length in the respective ions. In view of the extensive use which is made of D_2O as a solvent for the NMR study of complex molecules it is desirable that the origin of these effects should be identified.

Acknowledgements

We wish to thank Professor D. W. A. Sharp for allowing access to the Ramalog spectrometer used to obtain the above data and Mr. F. Mackie for assistance with obtaining spectra.

References

- 1 R. H. Nuttall and D. M. Stalker, *J. Chem. Soc. Dalton*, 1884 (1977).
- 2 Y. Fujiwara and C. N. Reilley, *Anal. Chem.*, **40**, 890 (1968).
- 3 M. Takeda, R. E. S. Ivazo, D. Garfinkel, I. H. Scheinberg and J. T. Edsall, *J. Am. Chem. Soc.*, **80**, 3813 (1958).
- 4 S. A. S. Ghazanhafer, J. T. Edsall and D. V. Myers, *ibid.*, **86**, 559 (1964).
- 5 T. L. James and J. H. Noggle, *ibid.*, **91**, 3424 (1969).
- 6 D. T. Sawyer and J. E. Tackett, *ibid.*, **85**, 314 (1963).
- 7 K. Nakamoto, Y. Morimoto and A. E. Martell, *ibid.*, **85**, 309 (1963).
- 8 K. Krishnan and R. A. Plane, *ibid.*, **90**, 3195 (1968).
- 9 J. B. Terrill and C. N. Reilley, *Anal. Chem.*, **38**, 1876 (1966).
- 10 M. Paabo and R. G. Bates, *Anal. Chem.*, **41**, 283 (1969).